dynamic processes in cells (*a systems approach to biology*)

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quantitative to qualitative dynamics



$$\frac{dx}{dt} = k_1 - k_2 x$$

from a quantitative analytical solution

$$x(t) = \frac{k_1}{k_2} + \left(x(0) - \frac{k_1}{k_2}\right) \exp(-k_2 t)$$





nonlinear qualitative dynamics

$$\frac{dx}{dt} = k_1 x (1-x) - k_2 x$$



bifurcation – change in parameter values that gives rise to a qualitative change of the dynamics in the state space

state space "landscapes" and parameter "geography"



nonlinear dynamics can be very complex

the dynamics may never reach a steady state. a trajectory may exhibit "sensitive dependence to initial conditions" ("chaos") but remain within a bounded region of state space, giving rise to a "strange attractor"

$$\frac{dx}{dt} = p_0 + y - z(p_1 + y)$$
$$\frac{dy}{dt} = p_2 + z - x(p_3 + z)$$
$$\frac{dz}{dt} = p_4 + x - y(p_5 + x)$$





"Does the flap of a butterfly's wings in Brazil set off a tornado in Texas?"

1917-2008

http://www.chaoscope.org/index.htm Olsen, Degn, *"Chaos in biological systems"*, Quart Rev Biophys **18**:165-225 1985 Scott, **Chemical Chaos**, OUP 1993



state space landscapes and cellular identity

Conrad Waddington

Sui Huang

Rudolf Jaenisch



Slack, "Conrad Hal Waddington: the last renaissance biologist?", Nature Rev Genetics **3**:889-95 2002. Huang, "Reprogramming cell fates: reconciling rarity with robustness", Bioessays, **31**:546-60 2009; Hana, Saha, Jaenisch, "Pluripotency and cellular reprogramming: facts, hypotheses and unresolved issues", Cell **143**:508-25 2010.

one gene, two states

positive feedback by a gene on itself is a potential way to create bistability



the first thing to calculate are the **steady states** of the dynamical system because they are the skeleton around which the dynamics takes place

method of nullclines (2D systems only)



but what is the stability of these steady states?

linearisation (lecture 4-16)

nonlinear system



nonlinear dynamical system

$$\frac{dx_1}{dt} = f_1(x_1, x_2, \cdots, x_n; k_1, \cdots, k_m)$$

$$\frac{dx_2}{dt} = f_2(x_1, x_2, \cdots, x_n; k_1, \cdots, k_m)$$

$$\vdots \quad \vdots \quad \vdots$$

$$\frac{dx_n}{dt} = f_n(x_1, x_2, \cdots, x_n; k_1, \cdots, k_m)$$



linear system

linear system

$$\frac{dy}{dt} = A.y \quad y = x - x_*$$
$$A = \left(\frac{\partial f_i}{\partial x_j}\right)\Big|_{x = x_*}$$

Jacobian matrix of partial derivatives

stability of steady states (lecture 4-4)



the matrix A has the eigenvalue λ with eigenvector x if $Ax = \lambda x$

 $\det(A - \lambda I) = 0$

a nonlinear dynamical system is stable at a steady state if all the eigenvalues of its Jacobian matrix, evaluated at the steady state, have negative real parts

stability theorem for genetic auto-regulation

assume general transcription & translation functions, linear degradation and arbitrary (positive or negative) feedback



$$\frac{dx_1}{dt} = f(x_2) - ax_1$$

$$\frac{dx_2}{dt} = g(x_1) - bx_2$$

a, b > 0

nullcline geometry determines stability

 x_1 nullcline, in the 1st quadrant, crosses above x_2 nullcline, in the 1st or 4th quadrants



 x_1 nullcline, in the 1st quadrant, crosses below x_2 nullcline, in the 1st quadrant



see the "nullcline theorem" handout for details

steady-state stability for positive feedback





how do we make **bistability** with the "off" state and the "on" state both stable?

bistability requires cooperativity ("sharpness")



positive feedback has to be combined with a sigmoidal ("S-shaped") nullcline.

this is "cooperativity" or "sharpness"

a common way to introduce cooperativity is to assume some kind of Hill function





hill functions for bistability

transcriptional priming



Laszlo et al, "Multilineage transcriptional priming and determination of alternate hematopoietic cell fates", Cell **126**:755-66 2006

hill functions for excitability



Kalmar, Lim, Hayward, Munoz-Descalzo, Nichols, Garcia-Ojalvo, Martinez-Arias, "Regulated fluctuations in Nanog expression mediate cell fate decisions in embryonic stem cells", PLoS Biol **7**:e1000149 2009

the hill function fits data on sharp gene expression



"consistent with the idea that Hb transcription is activated pairwise by cooperative binding of effectively five Bcd molecules" cooperativity

it is widely assumed that gene regulatory sharpness corresponding to a Hill coefficient of \mathbf{k} can be obtained from $\mathbf{n} = \mathbf{k}$ binding sites

Gregor, Tank, Wieschaus, Bialek, "Probing the limits to positional information", Cell **130**:153-64 2007

but the hill function lacks justification

the Hill function was introduced to fit data on oxygen binding to haemoglobin

 $\frac{x^h}{1+x^h}$



A V Hill, "The combinations of haemoglobin with oxygen and with carbon monoxide", Biochem J **7**:471-80 1913

Archibald Vivian Hill 1886 - 1977

"The Hill equation remains what Hill intended it to be: an empirical descriptor"

Engel, "A hundred years of the Hill equation", Biochem J 2013

"Despite its appealing simplicity, the Hill equation is not a physically realistic reaction scheme, raising the question of whether it should be abandoned in favor of realistic schemes; at the very least, its limitations should be more widely recognized"

Weiss, "The Hill equation revisited: uses and misuses", FASEB J 11:835-41 1997

and is not a valid GRF



the degree n is the number of sites and the coefficient c_k in the GRF is proportional to the steady-state probability of k sites out of n being occupied

this makes no physical sense for the Hill function ... even when the coefficient ${m h}$ is an integer

$$\frac{x^h}{1+x^h}$$

sharpness in gene regulation

$$f_n(x) = \frac{c_n x^n}{1 + c_1 x + \dots + c_n x^n} \qquad \begin{array}{c} \text{equilibrium GRF for} \\ \text{a single TF} \end{array}$$

measure sharpness of the gene regulation function using two properties – "steepness" and "position"



Hill functions are GRFs



Estrada, Wong, DePace, Gunawardena, "Information integration and energy expenditure in gene regulation", Cell **166**:234-44 2016